

CSF-1自背根节向脊髓转运对长春新碱诱导神经病理性痛大鼠小胶质细胞活化及其炎症的影响

付宝军¹, 姜静静¹, 黄玉琼¹, 林宗航¹, 李恒¹

(1. 广州医科大学附属第六医院//清远市人民医院麻醉科, 广东 清远 511518)

摘要:【目的】探讨CSF-1自背根节向脊髓转运对长春新碱诱导神经病理性痛大鼠小胶质细胞活化的影响。【方法】鞘内置管成功雄性SD大鼠54只,体质量200~230 g,10~12周龄,采用随机数字表法分为3组(每组18只):对照组(Control)、化疗诱发神经病理性痛+鞘内注射正常IgG组(CINP)、化疗诱发神经病理性痛+鞘内注射CSF1中和抗体(CINP+anti)。隔日腹腔注射长春新碱125 μg/kg(共计4次)建立CINP动物模型。分别采用机械缩足反射阈值(MWT)和热缩足反射潜伏期(TWL)评价大鼠机械痛敏和热痛敏;Western blotting法和免疫荧光化学检测CSF-1以及小胶质细胞标志物Iba1表达;RT-PCR法检测CSF-1 mRNA和Iba1 mRNA表达;ELISA法检测TNF-α、IL-6和IL-1β。【结果】与Control组比较,CINP组大鼠在长春新碱首次注射后第3、5、7天MWT和TWL明显降低($P<0.001$);与CINP组比较,CINP+anti组MWT和TWL在第5、7天明显升高($P<0.001$);与Control组比较,CINP组背根节CSF-1蛋白及mRNA表达、脊髓Iba1mRNA表达、脊髓CSF-1和Iba1蛋白表达、脊髓Iba1和背根节CSF-1荧光强度、脊髓TNF-α、IL-6和IL-1β表达明显上调($P<0.01, P<0.001$);与CINP组比较,CINP+anti组背根节和脊髓CSF-1蛋白表达、脊髓Iba1蛋白及mRNA表达、脊髓Iba1和背根节CSF-1荧光强度、脊髓TNF-α、IL-6和IL-1β表达明显下调($P<0.05, P<0.01, P<0.001$)。【结论】CSF-1自背根节向脊髓转运参与长春新碱诱导神经病理性疼痛发生过程,其机制可能与大鼠脊髓小胶质细胞活化及其炎症反应有关。

关键词:长春新碱;神经病理性痛;小胶质细胞;集落刺激因子-1

中图分类号:R338.1

文献标志码:A

文章编号:1672-3554(2020)05-0707-09

Effects of CSF-1 Transport from Dorsal Root Ganglion to Spinal Cord on Activation of Microglia and Inflammation in Rats with Neuropathic Pain Induced by Vincristine

FU Bao-Jun¹, JIANG Jing-Jing¹, HUANG Yu-Qiong¹, LIN Zhong-Hang¹, LI Heng¹

(1. Department of Anesthesiology, The Sixth Affiliated Hospital of Guangzhou Medical University// Qingyuan People Hospital, Qingyuan 511518, China)

Correspondence to: FU Bao-Jun; E-mail: fubaojun2004@126.com

Abstract: 【Objective】 To investigate the effects of CSF-1 transport from dorsal root ganglion to spinal cord on activation of microglia in rats with neuropathic pain induced by vincristine. 【Methods】 A total of 54 male 10-12-week old SD rats with successful intrathecal catheterization, weighing 200-230 g, were divided into three groups according to the random number table method ($n=18$): Control group (Control), Chemotherapy-induced Neuropathic Pain+intrathecal injection of IgG group (CINP), Chemotherapy-induced neuropathic pain+intrathecal injection of CSF-1 neutralizing antibody (CINP+anti). The animal model of CINP was established by intraperitoneal injection of vincristine 125 μg/kg on four alternate days. Mechanical allodynia and heat hyperalgesia were evaluated by MWT and TWL, respectively. The expression of CSF-1 and microglial marker Iba1 were detected by immunofluorescence chemistry and Western blotting. The mRNA expression of CSF-1 and Iba1 was measured by RT-PCR. The expression TNF-α, IL-6 and IL-1β were

收稿日期:2020-03-26

基金项目:广东省医学科研基金(A2019050);清远市科技计划项目(2018B066)

作者简介:付宝军,通信作者,副主任医师,研究方向:神经病理性疼痛,E-mail: fubaojun2004@126.com

determined by ELISA. 【Results】 Compared with Control group, MWT and TWL in C1NP group decreased significantly on the 3rd, 5th and 7th day after the first injection of vincristine ($P<0.01$), MWT and TWL in C1NP+anti group increased significantly on the 5th and 7th day compared with C1NP group ($P<0.01$). Compared with Control group, the protein and mRNA expression of CSF-1 in DRG, the mRNA expression of Iba1 in spinal cord, the protein expression of CSF-1 and Iba1 in spinal cord, the immunofluorescence intensity of CSF-1 in DRG and Iba1 in spinal cord, and the expression of TNF- α , IL-6 and IL-1 β in spinal cord were significantly up-regulated in C1NP group ($P<0.01$, $P<0.001$). Compared with C1NP group, the protein expression of CSF-1 in DRG and spinal cord, the protein and mRNA expression of Iba1 in spinal cord, the immunofluorescence intensity of spinal Iba1 and CSF-1 in DRG, and the expression of TNF- α , IL-6 and IL-1 β were obviously downregulated in C1NP+anti group ($P<0.05$, $P<0.01$, $P<0.001$). 【Conclusion】 The transport of CSF-1 from dorsal root ganglion to spinal cord is involved in the process of neuropathic pain induced by vincristine, and its mechanism may be related to the activation of microglia and inflammatory reaction in rat spinal cord.

Key word: vincristine; neuropathic pain; microglia; colony stimulating factor-1

[J SUN Yat-sen Univ (Med Sci), 2020, 41 (5): 707-715]

长春新碱是一种常用化疗药物,被用于治疗各种癌症,特别是急性淋巴细胞白血病、霍奇金淋巴瘤和非霍奇金淋巴瘤,但它常常诱导产生神经病理性疼痛(chemotherapy-induced neuropathic pain, C1NP)副作用,并呈剂量限制性^[1]。目前用于化疗引起的神经性疼痛的治疗药物仅限于抗惊厥药、阿片类药物和三环抗抑郁药^[2-3],但这些药物往往因其本身不可接受的副作用而限制临床应用。因此阐明C1NP发生机制,对于探索药物作用靶点、开发有效的治疗药物具有重大现实意义。集落刺激因子-1(colony stimulating factor-1, CSF-1)是一种巨噬细胞谱系特异性生长因子,在多种病理生理中发挥着作用,最近其在神经病理性疼痛中作用倍受关注^[4-6]近期研究证实^[7],Cre介导的感觉神经元内CSF1敲除可完全阻止神经损伤诱导的机械痛敏反应,减少小胶质细胞的活化和增殖。相反,鞘内注射CSF-1可诱导机械痛敏和小胶质细胞增生。另外,CSF-1介导关节炎促进关节炎导致痛觉过敏,以及阻断CSF-1治疗将有效预防和治疗关节炎导致痛觉过敏^[8-9],提示CSF-1在慢性疼痛信息调制中发挥重要作用,但是其在化疗药物诱导神经病理性疼痛中作用以及作用下游靶点未见报道。本研究拟观察长春新碱(vincristine)诱导神经病理性疼痛大鼠背根节和脊髓CSF-1的表达变化,并通过预先鞘内注射特异性CSF-1中和抗体,观察其对长春新碱导致的痛觉过敏大鼠脊髓小胶质细胞激活的影响和行为学的改变,探讨CSF-1在长春新碱诱导神经病理性疼痛中的作用及

其可能机制,为C1NP的发生机制提供了新的理论基础,同时也为C1NP的防治提供了新的科学依据。

1 材料与方法

1.1 实验动物与分组

清洁级雄性Sprague Dawley(SD)大鼠,体质量200~230 g,10~12周龄,由清远市人民医院实验动物中心提供[动物生产许可证号:SYXK(粤)2019-0206]。于12 h光照-黑暗循环、安静环境下单笼饲养,自由饮水摄食。室温(22.0 \pm 0.5) $^{\circ}$ C,湿度(55 \pm 10)%。所有实验操作均符合广州医科大学附属第六医院动物实验中心动物伦理要求并按照实验动物使用原则进行。鞘内置管成功SD大鼠54只,适应饲养1周后按随机数字表法分3组($n=18$):对照组(Control组);化疗痛组(C1NP组);化疗痛+CSF-1中和抗体组(C1NP+anti组)。CSF-1中和抗体(10 μ g 体积为10 μ L)(C1NP+anti组)或正常IgG(10 μ g 体积为10 μ L)(C1NP组和Control组)从第1天开始连续7 d于行为学测试前30 min鞘内给药,每次给药后用10 μ L生理盐水冲洗PE管;Control组为正常对照组,于鞘内给药后30 min腹腔注射等体积生理盐水,C1NP组、C1NP+anti组于鞘内给药后30 min腹腔注射长春新碱。

1.2 鞘内置管

参照Yaksh等^[10]报道方法鞘内置管,大鼠置管成功后观察其肢体运动情况,如有运动功能障碍

碍大鼠将被剔除后续实验。根据经导管注射利多卡因后30 s内出现双后肢麻痹现象确认导管位置正确,置管5 d后即可用于实验。

1.3 化疗痛动物模型建立

大鼠进行隔日腹腔注射长春新碱(浙江海正药业股份有限公司,中国),每次注射量为125 $\mu\text{g}/\text{kg}$ (共计4次),注射当天视为1 d,通过检测痛阈值变化确定化疗药物诱导的神经病理性疼痛模型建立成功。

1.4 行为学测定

参照Chaplan等^[11]报道的方法测定机械缩足反射阈值(mechanical withdrawal threshold, MWT),以up-down法推测阈值,并在阈值上下各刺激大鼠后肢足底中部5次,中位数法计算50%的反应阈值;参照Hargreaves等^[12]报道的方法测定热缩足反射潜伏期(thermal withdrawal latency, TWL)自动切断时间为20 s,以防止组织损伤。大鼠后肢足底中部热刺激强度在整个实验过程中保持一致。每只动物测定5次(间隔3 min),取其平均值为大鼠TWL值。在给药前、给药后第1、3、5、7天分别采用MWT和TWL评价大鼠机械痛敏和热痛敏。

1.5 Western blot实验

第7天,每组各取3只大鼠麻醉后断头处死,冰上取出L4-5脊髓节段及背根节,加入裂解液进行匀浆,4℃下12 000 r/min($r=5\text{ cm}$)离心5 min,并进行BCA蛋白定量后每份样品使用20 μg 蛋白质。配置分离胶和浓缩胶,当溴酚蓝染料前端电泳至分离胶末端处时即停止电泳,转膜后封闭2 h,加入抗 β -actin兔多克隆抗体(1:2 000, Abcam, 美国),抗CSF-1兔多克隆抗体(1:1 000, Abcam, 美国),抗Iba1(ionized calcium binding adapter molecule 1, Iba1)兔多克隆抗体(1:1 000, Abcam, 美国),4℃孵育过夜后洗膜3次(每次10 min)。加入HRP标记的山羊抗兔IgG(1:3 000, Abcam, 美国)室温孵育后洗膜、显色、曝光、显影,采用Image J软件检测目的蛋白条带及 β -actin蛋白条带的灰度值,目的蛋白条带灰度值/ β -actin蛋白条带灰度值比值作为目标蛋白表达量。

1.6 RT-qPCR检测

第7天,每组各取3只大鼠安乐死后进行检查。TRIzol[®] reagent(Invitrogen, 美国)提取大鼠L4-6脊髓及背根节总RNA,反转录为cDNA。用

2^{- $\Delta\Delta\text{CT}$} 法来测定Csf-1 mRNA、Iba1 mRNA含量。引物合成(北京天跟生化科技有限公司,中国)为:Csf-1上游引物:5'-TGCTAAGTGCTCTAGCCGAG-3';下游引物5'-CCCCAACAGTCAGCAAGAC-3'。上游引物:Iba1上游引物:5'-CCATGACCTTCCAA-GAGAATGC-3';下游引物:5'-ACCGGCTTGTGC-TGTAGTC。 β -actin上游引物:5'-CGTTGACATC-CGTAAAGACCTC-3';下游引物:5'-TAGGAGC-CAGGGCAGTAATCT-3'。扩增条件:94℃预变性5 min,94℃30 s,54℃30 s,72℃20 s,共45个循环,72℃延伸10 min。计算与内参照 β -actin的比值作为目的基因的相对表达量。

1.7 免疫组织荧光实验

第7天,每组大鼠取3只经腹腔注射1%戊巴比妥钠40 mg/kg麻醉后,用40 g/L多聚甲醛灌注固定,取大鼠腰4~5背根节及脊髓,40 g/L多聚甲醛后固定2 h,再先后于20%、30%蔗糖溶液中脱水,冰冻切片机切片,厚度14 μm 。免疫荧光染色:磷酸盐缓冲盐水(phosphate buffered saline, PBS)(0.01 mol/L, pH值7.4)洗片3次,20 min/次;加5%羊血清室温封闭2 h;加入抗Iba1及CSF-1兔多克隆抗体一抗(1:200, Cell Signaling Technology, 美国),4℃过夜孵育;次日复温至室温后PBS洗3次,15 min/次;分别加入Cy3标记的荧光二抗(1:1 000, Cell Signaling Technology, 美国),室温孵育2 h,PSB洗3次,20 min/次,晾干,封片剂封片;置荧光显微镜下拍照观察。

1.8 ELISA检测

第7天,L4~L5脊髓节段匀浆后于4℃,3 500 r/min($r=5\text{ cm}$)下离心10 min,提取上清液并在-80℃保存。按ELISA测定试剂盒说明书步骤进行操作。采用NK3酶标仪(GE公司,美国)于波长490 nm处测定光密度值,对应标准曲线得出TNF- α 、IL-6、IL-1 β 的含量。

1.9 统计学方法

符合正态分布的计量资料以均数 \pm 标准差表示,采用graphpad 7.0软件进行统计分析,行为学结果采用连续性重复测量方差分析,若组别和时间存在交互效应,进一步检验组间和时间点的单独效应,两两比较采用LSD- t 法;Western blotting、RT-PCR、免疫荧光化学、ELISA数据以单因素方差分析,两两比较采用Tukey's法; $P < 0.05$ 被认为有统计学差异。

2 结果

2.1 各组大鼠MWT和TWL变化情况

Control组、CINP组、CINP+anti组大鼠的行为学结果进行重复测量方差分析显示:MWT和TWL数据均满足球形分布($\chi^2 = 3.3, P = 0.92; \chi^2 = 13.42, P = 0.15$), 各组间差异有统计学意义($F = 24.69, P < 0.001; F = 21.25, P < 0.001$), 各组时间点差异有统计学意义($F = 37.86, P < 0.001; F = 40.15, P < 0.001$), 且分组和时间点有交互效应($F = 27.11, P < 0.001; F = 28.61, P < 0.001$)。两两比较分析比

较各组相同时间点MWT和TWL的差异, 结果表明各组基础痛觉阈值无统计学差异($F = 0.89, P = 0.37$), 而腹腔注射长春新碱后第3、5、7天, 大鼠的MWT[3 d: (12.60 ± 0.55) g; 5 d: (9.30 ± 0.84) g, 7 d: (8.10 ± 0.70) g, $P < 0.001$ vs. Control组]和TWL[3 d: (17.10 ± 0.55) s; 5 d: (15.18 ± 0.60) s; 7 d: (12.90 ± 0.74) s, $P < 0.001$ vs. Control组]均显著降低, 鞘内给予CSF1中和抗体明显增加化疗痛大鼠第5、7天MWT[5 d: (11.68 ± 1.10) g; 7 d: (10.60 ± 0.55) g, $P < 0.001$ vs. CINP组]和TWL[5 d: (17.18 ± 0.60) s; 7 d: (14.80 ± 0.50) s, $P < 0.001$ vs. CINP组; 图1]。

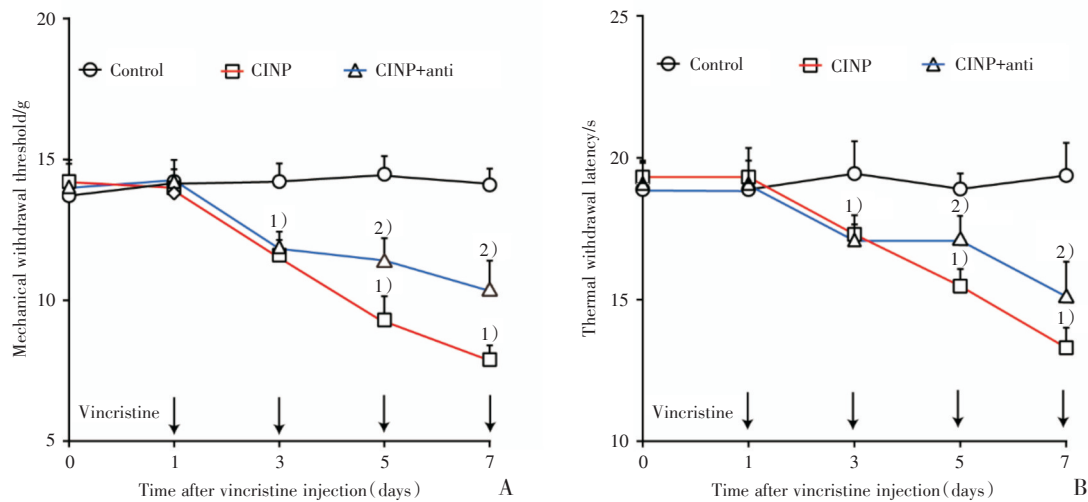


图1 各组大鼠MWT和TWL变化情况

Fig.1 MWT and TWL for rats in different groups

A: The time course of mechanical withdrawal threshold (MWT); B: The time course of thermal withdrawal latency (TWL). Pre-treatment with IgG does not produce analgesic effect in the CINP group. In contrast, pretreatment with CSF-1 neutralizing antibody significantly attenuates mechanical and thermal hyperalgesia following remifentanyl infusion at 5 d and 7 d. $F=37.86$ (A), $F=40.15$ (B). 1) $P < 0.001$, compared with the Control group; 2) $P < 0.001$, compared with the CINP group; $n=6$ per group.

2.2 各组大鼠背根节CSF-1蛋白及mRNA表达情况

长春新碱注射后第7天, 分别对Control组、CINP组、CINP+anti组大鼠背根节CSF-1蛋白及mRNA进行单因素方差分析结果显示差异有统计学意义(蛋白: $F = 129.40, P < 0.001$; mRNA: $F = 428.00, P < 0.001$); 两两比较结果显示CINP组大鼠背根节CSF-1蛋白及mRNA明显上调(CINP组: 蛋白ID = $0.32 \pm 0.02, P < 0.001$ vs. Control组; CINP组: mRNAID = $0.94 \pm 0.06, P < 0.001$ vs. Control组), 说明长春新碱腹腔注射导致CSF-1表达增加。鞘内给予CSF1中和抗体可以抑制长

春新碱诱导CSF-1蛋白及mRNA表达上调(CINP+anti组: 蛋白ID = $0.22 \pm 0.03, P < 0.01$ vs. CINP组; CINP+anti组: mRNAID = $2.02 \pm 0.06, P = 0.97$ vs. CINP组, 图2 A, B)。

2.3 各组大鼠背根节CSF-1免疫荧光表达情况

长春新碱注射后7天, 分别对Control组、CINP组、CINP+anti组大鼠背根节CSF-1荧光强度进行单因素方差分析结果显示差异有统计学意义($F = 114.20, P < 0.001$); 两两比较结果显示CINP组大鼠背根节CSF-1荧光强度明显增强(CINP组: $252.74 \pm 33.25, P < 0.001$ vs. Control组), 说明长春新碱腹腔注射导致背根节CSF-1表达增加。鞘内

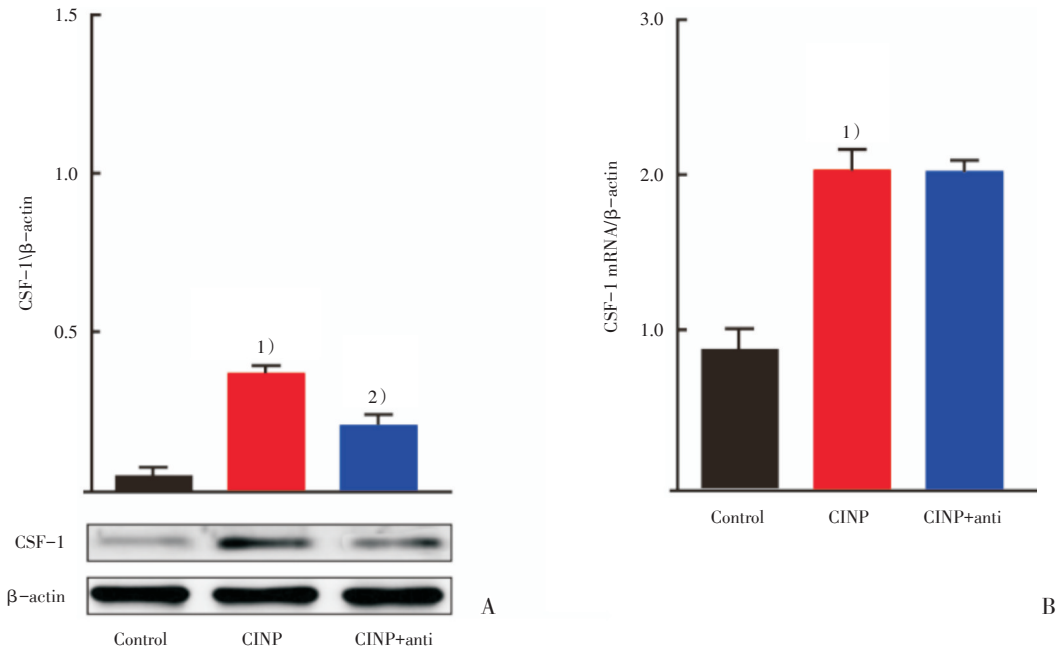


图2 各组大鼠背根节 CSF-1 蛋白及 mRNA 表达情况

Fig.2 protein and mRNA expression of CSF-1 in DRG of rats in different groups

A: Western blot shows that vincristine application increases the expression of CSF-1 protein in DRG. The effect was inhibited by intrathecal injection of CSF-1 neutralizing antibody. $F = 129.40$, 1) $P < 0.001$, 2) $P < 0.01$, $n = 3$ per group. B: PCR shows that vincristine application increases the expression of CSF-1 mRNA in DRG; the effect was not inhibited by intrathecal injection of CSF-1 neutralizing antibody. $F = 428.00$, 1) $P < 0.001$, $n = 3$ per group.

给予 CSF-1 中和抗体可以抑制长春新碱诱导 CSF-1 荧光强度增强 (C1NP+anti 组: 141.93 ± 12.21 , $P < 0.01$ vs. C1NP 组, 图 3 A~D)。

2.4 各组大鼠脊髓 CSF-1 蛋白及 mRNA 表达情况

长春新碱注射后第 7 天, 分别对 Control 组、C1NP 组、C1NP+anti 组大鼠脊髓 CSF-1 蛋白进行单因素方差分析结果显示差异有统计学意义 (蛋白: $F = 283.20$, $P < 0.001$; mRNA: $F = 0.94$, $P = 0.31$): 两两比较结果显示 C1NP 组大鼠脊髓 CSF-1 蛋白表达增强 (C1NP 组: 蛋白 $ID = 0.85 \pm 0.05$, $P < 0.001$

vs. Control 组), 说明腹腔注射长春新碱导致 CSF-1 表达增加。鞘内给予 CSF-1 中和抗体可以抑制长春新碱诱导脊髓 CSF-1 表达上调 (C1NP+anti 组: $ID = 0.25 \pm 0.05$, $P < 0.001$ vs. C1NP 组, 图 4 A~D)。

2.5 各组大鼠脊髓小胶质细胞标志物 Iba1 蛋白及 mRNA 表达情况

长春新碱注射后第 7 天, 分别对 Control 组、C1NP 组、C1NP+anti 组大鼠脊髓 Iba1 蛋白及 mRNA 进行单因素方差分析结果显示差异有统计学意义 (蛋白: $F = 70.63$, $P < 0.001$; mRNA: $F = 108.50$,

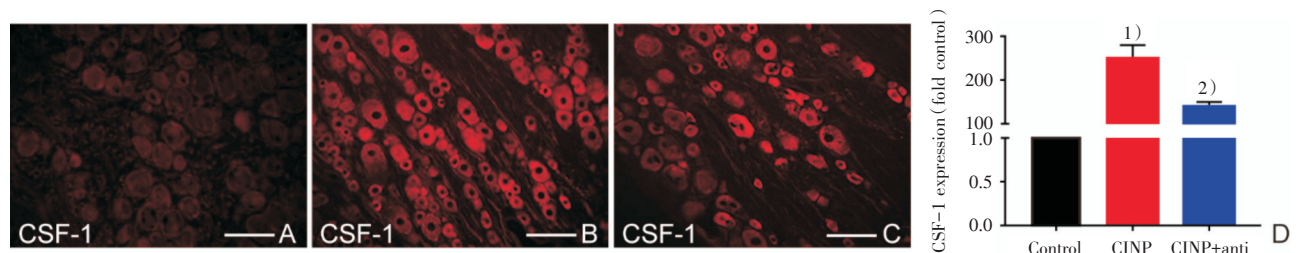


图3 各组大鼠背根节 CSF-1 免疫荧光表达情况

Fig.3 Immunofluorescence expression levels of CSF-1 in DRG of rats in different groups

A: Immunofluorescence of CSF-1 in the Control group; B: Immunofluorescence of CSF-1 in the C1NP group; C: Immunofluorescence of CSF-1 in the C1NP + anti group; D: The immunofluorescence relative density of CSF-1 in the Control, C1NP and C1NP+anti group at 7 d. $F = 114.20$. 1) $P < 0.001$, compared with the Control group; 2) $P < 0.01$, compared with the C1NP group; $n = 3$ per group. Scale bar: 50 μm

$P < 0.001$): 两两比较结果显示 CINP 组大鼠背根节 Iba1 蛋白及 mRNA 表达明显上调 [CINP 组(蛋白): 0.75 ± 0.08 , $P < 0.001$ vs. Control 组; CINP 组(mRNA): 0.95 ± 0.09 , $P < 0.001$ vs. Control 组], 说明长春新碱腹腔注射导致脊髓小胶质细胞活化。

鞘内给予 CSF-1 中和抗体可以抑制长春新碱诱导小胶质细胞活化 [CINP+anti 组(蛋白): 0.53 ± 0.06 , $P < 0.05$ vs. CINP 组; CINP+anti 组(mRNA): 0.63 ± 0.07 , $P < 0.01$ vs. CINP 组, 图 5 A, B]。

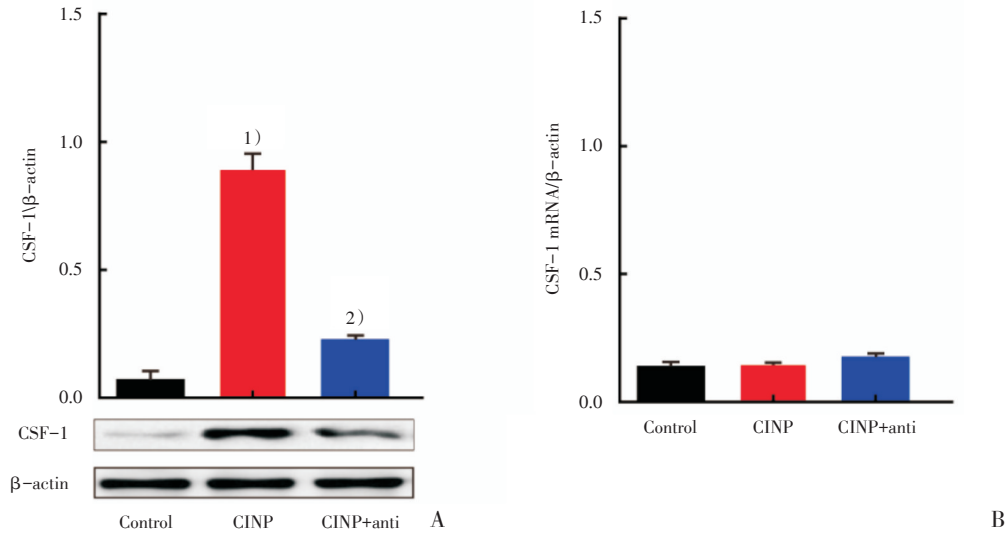


图 4 各组大鼠脊髓 CSF-1 蛋白及 mRNA 表达情况

Fig.4 Protein and mRNA expression of CSF-1 in spinal cord of rats in different groups

A: Western blot shows that vincristine application increased the expression of CSF-1 protein in spinal cord, and the effect was inhibited by intrathecal injection of CSF-1 neutralizing antibody. $F = 283.20$, 1) $P < 0.001$, compared with the Control group. 2) $P < 0.001$, compared with the CINP group, $n = 3$ per group. B: PCR shows that there was no significantly difference in the expression of CSF-1 mRNA in spinal cord among Control, CINP and CINP+anti groups. $F = 0.94$, $P = 0.31$, $n = 3$ per group.

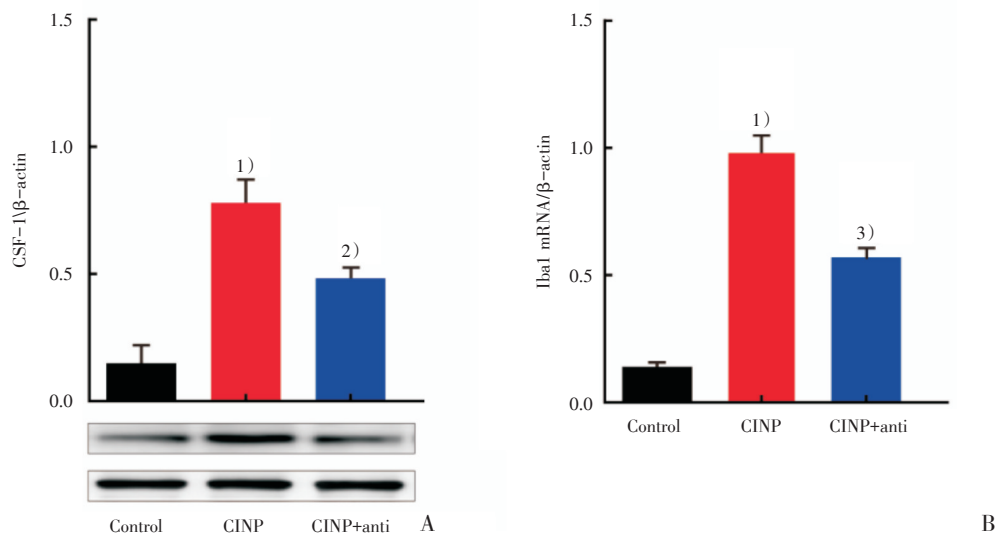


图 5 各组大鼠脊髓 Iba1 蛋白及 mRNA 表达情况

Fig.5 Protein and mRNA expression of Iba1 in spinal cord of rats in different groups

A: Western blot shows that vincristine application increased the expression of Iba1 protein in spinal cord, and the effect was inhibited by intrathecal injection of CSF-1 neutralizing antibody. $F = 70.63$, 1) $P < 0.001$, compared with the Control group; 2) $P < 0.05$, compared with the CINP group, $n = 3$ per group; B: PCR shows that vincristine application increased the expression of Iba1 mRNA in spinal cord; the effect was not inhibited by intrathecal injection of CSF-1 neutralizing antibody. $F = 108.50$, 1) $P < 0.001$, compared with the Control group; 3) $P < 0.01$, compared with the CINP group, $n = 3$ per group.

2.6 各组大鼠脊髓小胶质细胞标志物 Iba1 免疫荧光表达情况

长春新碱注射后第7天,分别对 Control 组、CINP 组、CINP+anti 组大鼠脊髓 Iba1 荧光强度进行单因素方差分析结果显示差异有统计学意义($F = 91.47, P < 0.001$);两两比较结果显示 CINP 组大鼠脊髓 Iba1 荧光强度增强(CINP 组: $20.02 \pm 2.50, P < 0.001$ vs. Control 组),说明长春新碱腹腔注射导致脊髓小胶质细胞活化。鞘内给予 CSF1 中和抗体可以抑制长春新碱诱导 Iba1 荧光强度增强(CINP + anti 组: $6.01 \pm 1.82, P < 0.001$ vs. CINP 组,图 6 A~D),提示 CSF-1 作用发挥依赖于脊髓小胶质细胞。

2.7 各组大鼠脊髓 TNF- α 、IL-6 和 IL-1 β 表达情况

长春新碱注射后第7天,分别对 Control 组、

CINP 组、CINP+anti 组大鼠脊髓 TNF- α 、IL-6 和 IL-1 β 表达进行单因素方差分析结果显示差异有统计学意义($F = 20.79, P < 0.01; F = 20.81, P < 0.01; F = 23.23, P < 0.01$);两两比较结果显示,与 Control 组比较,CINP 组大鼠脊髓 TNF- α 、IL-6 和 IL-1 β 表达增加[(251.45 ± 22.14)pg/mg, $P < 0.01$, vs. Control 组; (215.42 ± 37.19)pg/mg, $P < 0.01$, vs. Control 组; (168.43 ± 25.69)pg/mg, $P < 0.01$, vs. Control 组],说明长春新碱腹腔注射导致脊髓炎症反应。鞘内给予 CSF1 中和抗体可以抑制长春新碱诱导 TNF- α 、IL-6 和 IL-1 β 表达上调[(165.48 ± 34.26)pg/mg, $P < 0.05$, vs. CINP 组; (138.71 ± 16.22)pg/mg, $P < 0.05$, vs. CINP 组; (100.23 ± 18.29)pg/mg, $P < 0.05$, vs. CINP 组,图 7 A~C)]。

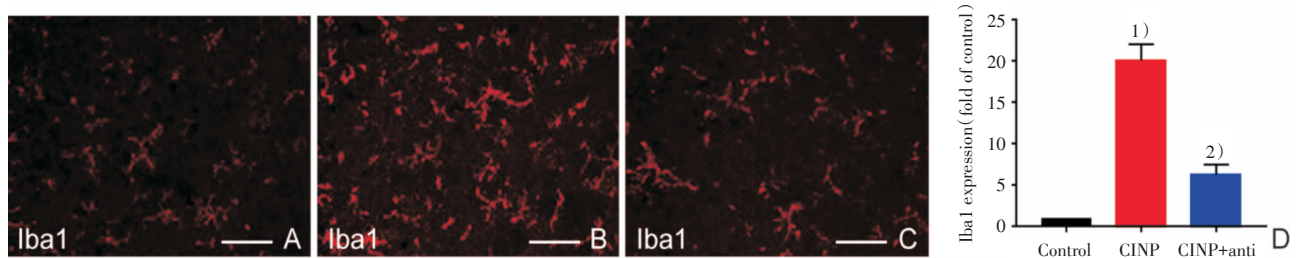


图 6 各组大鼠脊髓 Iba1 免疫荧光表达情况

Fig.6 Immunofluorescence expression levels of Iba1 in spinal cord of rats in different groups

A: Immunofluorescence of spinal Iba1 in the Control group; B: Immunofluorescence of spinal Iba1 in the CINP group; C: Immunofluorescence of spinal Iba1 in the CINP + anti group. D: The immunofluorescence relative density of CSF-1 in the Control, CINP and CINP + anti group at 7 d. $F = 91.47$. 1) $P < 0.001$, compared with the Control group; 2) $P < 0.001$, compared with the CINP group, $n = 3$ per group; Scale bar: 50 μm

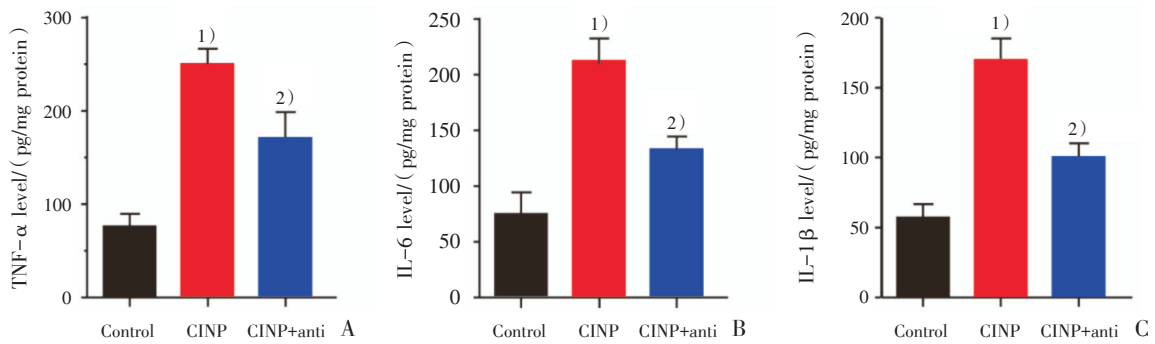


图 7 各组大鼠脊髓 TNF- α 、IL-6 和 IL-1 β 表达情况

Fig.7 expression levels of TNF- α , IL-6 and IL-1 β in spinal cord of rats in different groups

A: ELISA shows that vincristine application increased the expression of TNF- α protein in spinal cord; the effect was inhibited by intrathecal injection of CSF-1 neutralizing antibody. $F = 20.79$, 1) $P < 0.01$, compared with the Control group; 2) $P < 0.05$, compared with the CINP group, $n = 3$ per group; B: ELISA shows that vincristine application increased the expression of IL-6 protein in spinal cord; the effect was inhibited by intrathecal injection of CSF-1 neutralizing antibody. $F = 20.81$, 1) $P < 0.01$, compared with the Control group; 2) $P < 0.05$, compared with the CINP group, $n = 3$ per group. C: ELISA shows that vincristine application increased the expression of IL-1 β protein in spinal cord; the effect was inhibited by intrathecal injection of CSF-1 neutralizing antibody. $F = 20.81$, 1) $P < 0.01$, compared with the Control group; 2) $P < 0.05$, compared with the CINP group, $n = 3$ per group.

3 讨论

本研究参照许爱军等^[13]方法建立 C1NP 大鼠模型,该模型由于可操作性强、重复性好、较好模拟临床 C1NP 特征等优点,已广泛应用于验证药物和探讨机制的动物实验中。长春新碱首次给药后 3 d,大鼠 MWT 和 TWL 开始明显降低,并伴有跛行、抬足、舔舐足底增多以及自发嘶叫等疼痛行为学变化,并持续到本实验观察结束,表明 C1NP 模型制备成功。我们团队前期研究证实,长春新碱诱导神经病理性疼痛形成过程中,脊髓小胶质细胞活化介导 C1NP 大鼠痛觉敏化过程,但是小胶质细胞活化调节机制仍不清楚。

CSF-1 是一种细胞因子,通过与 III 型受体酪氨酸激酶偶联 CSF-1 受体结合在调节单核细胞、巨噬细胞、小胶质细胞存活、增殖和分化中发挥重要作用^[14]。随着神经病理性疼痛机制研究不断深入,细胞因子,尤其 CSF-1,备受广泛关注,越来越多证据显示 CSF-1 在神经病理性疼痛中发挥重要作用^[4-5],结果表明,与 Control 组比较,C1NP 组大鼠脊髓 CSF-1 蛋白表达明显上调,而 CSF-1 mRNA 表达差异无统计学意义,提示 CSF-1 原发产生部位并非在脊髓,可能存在其他部位 CSF-1 蛋白向脊髓转运;背根节免疫荧光结果表明,与 Control 组比较,C1NP 组大鼠背根节 CSF-1 荧光表达明显上调,脊髓 CSF-1 蛋白表达上调是否源于背根节 CSF-1 轴向转运,前期实验通过在构建 C1NP 模型前结扎 L4/L5 背根,结果脊髓 CSF-1 表达与正常对照组差异无统计学意义,进一步证实背根节 CSF-1 向脊髓转运。与我们研究不一致,另有研究^[15]证实大鼠缺血痛模型建立后 6 h 脊髓背角活化的星形胶质细胞产生 CSF-1,相反,星形胶质细胞抑制剂氟代柠檬酸显著抑制缺血 6 h 后 CSF-1 上调,提示 CSF-1 其产生部位在脊髓,导致上述实验 CSF-1 产生部位差异可能与使用模型以及动物种属不同有关。

既然本研究提示可能存在 CSF-1 向脊髓转

运,那么 CSF-1 对长春新碱诱发痛觉过敏调节机制如何?最近的研究表明^[7],脊神经结扎大鼠背根节初级感觉神经元中 CSF-1 表达上调,通过轴向转运作用于脊髓小胶质细胞表面 CSF-1 受体,进而激活小胶质细胞。与上述研究一致的是,本研究腹腔注射长春新碱后,C1NP 组脊髓 Iba1(小胶质细胞标志物)蛋白和 mRNA 表达显著升高,相反,鞘内注射 CSF-1 中和抗体明显抑制 C1NP 大鼠脊髓 Iba1 蛋白和 mRNA,提示,通过鞘内注射 CSF-1 中和抗体阻断 CSF-1 信号明显抑制脊髓小胶质细胞活化,CSF-1 信号作用是通过脊髓小胶质细胞活化实现的。

为了进一步证实 CSF-1 信号调控脊髓小胶质细胞活化,我们采用脊髓 Iba1 免疫荧光化学从形态学角度探讨阻断 CSF-1 信号对小胶质细胞活化的影响,结果显示鞘内注射 CSF-1 中和抗体阻断 CSF-1 信号明显抑制脊髓小胶质细胞活化。一般认为,胶质细胞介导神经炎症在慢性疼痛中枢敏化中发挥作用^[16-17],活化胶质细胞释放促炎细胞因子,如:TNF- α 、IL-6 和 IL-1 β ,其与感觉神经元表面受体结合,增强神经元兴奋性,促发痛觉过敏^[18-19],本研究表明:腹腔注射长春新碱后,C1NP 组大鼠脊髓 TNF- α 、IL-6 和 IL-1 β 表达显著上调,相反,鞘内注射 CSF-1 中和抗体明显抑制 C1NP 大鼠脊髓 TNF- α 、IL-6 和 IL-1 β 表达,提示,通过鞘内注射 CSF-1 中和抗体阻断 CSF-1 信号明显抑制脊髓神经炎症,即 CSF-1 信号作用发挥是依赖于神经炎症。

最近的研究显示,在骨肿瘤诱导的疼痛动物模型中,背根节 CSF-1 介导痛觉过敏依赖于钠通道转录水平调控^[5],而后者已被证实在多种炎症及神经病理性疼痛中发挥着重要作用^[20-21]。因此,背根节水平的 CSF-1 在化疗药物引起的痛觉过敏中的具体作用和机制有待研究进一步证实。

总之,CSF-1 自背根节向脊髓转运参与长春新碱诱导神经病理性疼痛发生过程,其机制可能与大鼠脊髓小胶质细胞活化及其炎症反应有关。

参考文献

[1] Brewer JR, Morrison G, Dolan ME, et al. Chemo-

therapy-induced peripheral neuropathy: current status and progress[J]. *Gynecol Oncol*, 2016, 140(1): 176-183.

- [2] Busse JW, Wang L, Kamaleldin M, et al. Opioids for chronic noncancer pain: a systematic review and meta-analysis[J]. *JAMA*, 2018, 320(23): 2448-2460.
- [3] Fornasari D. Pharmacotherapy for neuropathic pain: a review[J]. *Pain Ther*, 2017, 6(Suppl 1): 25-33.
- [4] Zhou LJ, Peng J, Xu YN, et al. Microglia are indispensable for synaptic plasticity in the spinal dorsal horn and chronic pain[J]. *Cell Rep*, 2019, 27(13): 3844-3859.
- [5] Zhang F, Wang Y, Liu Y, et al. Transcriptional regulation of voltage-gated sodium channels contributes to GM-CSF-induced pain[J]. *J Neurosci*, 2019, 39(26): 5222-5233.
- [6] Boakye PA, Rancic V, Whitlock KH, et al. Receptor dependence of BDNF actions in superficial dorsal horn: relation to central sensitization and actions of macrophage colony stimulating factor 1[J]. *J Neurophysiol*, 2019, 121(6): 2308-2322.
- [7] Guan Z, Kuhn JA, Wang X, et al. Injured sensory neuron-derived CSF1 induces microglial proliferation and DAP12-dependent pain[J]. *Nat Neurosci*, 2016, 19(1): 94-101.
- [8] Conaghan PG, Cook AD, Hamilton JA, et al. Therapeutic options for targeting inflammatory osteoarthritis pain[J]. *Nat Rev Rheumatol*, 2019, 15(6): 355-363.
- [9] Saleh R, Lee MC, Khiew SH, et al. CSF-1 in inflammatory and arthritic pain development[J]. *J Immunol*, 2018, 201(7): 2042-2053.
- [10] Yaksh TL, Rudy TA. Chronic catheterization of the spinal subarachnoid space[J]. *Physiol Behav*, 1976, 17(6): 1031-1036.
- [11] Chaplan SR, Bach FW, Pogrel JW, et al. Quantitative assessment of tactile allodynia in the rat paw[J]. *J Neurosci Method*, 1994, 53(1): 55-63.
- [12] Hargreaves K, Dubner R, Brown F, et al. A new and sensitive method for measuring thermal nociception in cutaneous hyperalgesia[J]. *Pain*, 1988, 32(1): 77-88.
- [13] 许爱军,曹菲,田玉科.长春新碱诱发外周神经病理性疼痛模型的建立[J].*中国疼痛医学杂志*, 2008, 14(3):163-166.
- Xu AJ, Cao F, Tian YK. Study on the rat model of vincristine-induced peripheral neuropathic pain[J]. *Chin J Pain Med*, 2008, 14(3): 163-166.
- [14] Dhagat U, Hercus TR, Broughton SE, et al. The mechanism of GM-CSF inhibition by human GM-CSF auto-antibodies suggests novel therapeutic opportunities[J]. *Mabs*, 2018, 10(7):1018-1029.
- [15] Tang Y, Liu L, Xu D, et al. Interaction between astrocytic colony stimulating factor and its receptor on microglia mediates central sensitization and behavioral hypersensitivity in chronic post ischemic pain model[J]. *Brain Behav Immun*, 2018, 68:248-260. doi: 10.1016/j.bbi.2017.10.023
- [16] Doan RA, Monk KR. Glia in the skin activate pain responses[J]. *Science*, 2019, 365(6454): 641-642.
- [17] Ji RR, Donnelly CR, Nedergaard M. Astrocytes in chronic pain and itch[J]. *Nat Rev Neurosci*, 2019, 20(11):667-685.
- [18] 李秋月,许海玉,杨洪军.促炎因子TNF- α , IL-1 β , IL-6在神经病理性疼痛中的研究进展[J].*中国中药杂志*, 2017, 42(19):3709-3712.
- Li QY, Xu HY, Yang HJ. Effect of proinflammatory factors TNF- α , IL-1 β , IL-6 on neuropathic pain[J]. *Chin J Chin Mater Med*, 2017, 42(19): 3709-3712.
- [19] Chen J, Cong X, Zhan X, et al. Effects of Parecoxib on pain threshold and inflammatory factors IL-1 β , IL-6 and TNF- α in spinal cord of rats with bone cancer pain[J]. *J Coll Physicians Surg Pak*, 2019, 29(6):528-531.
- [20] Urru M, Muzzi M, Coppi E et al. Dextrampipexole blocks Nav1.8 sodium channels and provides analgesia in multiple nociceptive and neuropathic pain models[J]. *Pain*, 2020, 16(4): 831-841.
- [21] Lee JY, Kam YL, Oh J, et al. HYP-17, a novel voltage-gated sodium channel blocker, relieves inflammatory and neuropathic pain in rats[J]. *Pharmacol Biochem Behav*, 2017, 153:116-129. doi:10.1016/j.pbb.2016.12.013

(编辑 孙慧兰)